SUBJECT/COURSE SYLLABUS

1.	Course name in Polish and English		
	Animal Cell Culture Techniques		
	Hodowle komórek i tkanek zwierzęcych		
2.	Scientific discipline		
	Medical sciences		
	Biotechnology		
3.	Language of instruction		
	English		
4.	Unit conducting the course		
	Faculty of Biotechnology		
5.	Type of course		
	elective - choice limited to Animal Cell Culture Techniques and Plant Cell		
	Culture Techniques		
6.	Field of study		
	Biotechnology		
7.	Level of study		
	first-cycle		
8.	Year of study		
	2nd		
9.	Semester		
	summer		
10.	Course form and number of hours		
	Lecture: 15 h		
	Laboratory: 30 h		
11.	Prerequisites in terms of knowledge, skills, and social competencies for the		
	course:		
	basic knowledge about biology and biochemistry in the first years of		
	bachelor studies, and basic skills in laboratory work .		
12.	Learning objectives for the course:		
	LECTURE: Acquiring knowledge about work with animal cell and tissue cultures.		
	LABORATORY: Students will gain knowledge of culturing animal cells. They will		
	learn how to culture animal cells derived from normal tissues and cancer cell		
	lines. General knowledge of fluorescence/confocal microscopy, including its		
	applications, will be passed to the Students. Students will also gain an		
	understanding of quantitative methods used in the study of animal cells,		
	including the assessment of cell viability and cell cycle distribution under drug treatment.		
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13. |Curriculum content:

LECTURE:

Attending the lecture will allow the students to get familiar with specificity of working in the laboratory where animal cells are cultured. During the lectures, the following topics are mentioned and discussed:

- safety of work with animal cell and tissue cultures, and genetically modified microorganisms;
- organization of animal cell culture laboratory;
- preservation of sterile conditions during work with animal cells;
- cells' authentication;
- composition and types of the most commonly used media in animal cell cultures;
- sources of cells and tissues;
- characterization of primary cell cultures and cell lines;
- examples of modern applications employing animal cell cultures in research aiming to solve scientific problems raised in biology, pharmacology, biotechnology, and medicine.

LABORATORY:

Active participation in laboratory classes will allow students to become familiar with the specifics of working in a laboratory where animal cells are cultured. During the practical course, students will learn to culture primary cells and cancer cell lines, examine cell viability, count cells, and subculture them. Students will also gain skills in preparing slides with fixed and stained cells and in analyzing cell morphology and the distribution of selected actin cytoskeleton proteins within a cell using an inverted light microscope and a confocal microscope, respectively. Some quantitative analyses, such as estimating cell viability and cell cycle distribution in the tested cells, are also done.

14.	Description	of learning	outcomes
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Student:

- has knowledge of the basic techniques and research tools used for/in animal cell cultures;
- knows how to combine theoretical knowledge of mammalian cell cultures with its practical application in research;
- is familiar with and implements the basic principles of health and safety and ergonomics procedures in the laboratory, knows and follows procedures of work with genetically modified microorganisms;
- applies basic physicochemical and biochemical techniques necessary for studying biological processes (including processes taking place in cultured cells);
- has skills in culturing cells of higher organisms;

Symbols for relevant directional learning outcomes:

K1_W08

K1_W09

K1_W10, K1_K05

K1 U01

K1_U02

	sources, uses professional scientific language and terminology specific to the course topics; • performs standard animal cell biology experiments safely using proper laboratory	K1_U03, K1_U04, K1_U09 K1_U05			
	analyzes experimental data and describes the results accurately and objectively:	K1_U05, K1_U06 K1_U13, K1_K03			
	 planning and organizing work, as well as addressing a scientific problem; understands the need for updating knowledge and skills in the field of biotechnology. 	K1_K01			
15.	Mandatory literature:				
	 laboratory manual provided by the teachers. 				
	Recommended literature:				
	 R.I. Freshney, Culture of Animal cells - a manu and specialized applications, Willey-Blackwell; other sources provided/indicated by the teachers. 	al of basic techniques			
	other sources provided/indicated by the teachers.				
16.	omes:				
	 LABORATORY: Students will be assessed on their ability to safely and competently execute laboratory procedures and manage their time efficiently during all scheduled lab sessions. Active participation and attendance are mandatory. 				
	• Final written test.				
	LECTURE:				
	 final written test (the condition for taking the test is to obtain a post grade in the laboratory classes). 				
17.	Conditions and form of credit for individual component	ts of the course:			
	LABORATORY:				
	 mandatory presence and active participation in the classes; positive test result. 				
	LECTURE: • positive test result.				
18.	Student workload expressed in teaching hours and EC credits	TS number of hours allocated for the course of a given type of classes			
10.	Classes (according to the study plan) with the				
	instructor:laboratory (including an introduction lecture)lecture	30 h 15 h			
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Student's own work (including participation in work), e.g.: • preparation for the classes • reading the manual and literature indices • preparation for the final tests	50 h
Total number of class hours:	95 h
Number of ECTS credits: laboratorylecture	2 ECTS 2 ECTS